Data Issues of Using Matching Methods to Estimate Treatment Effects:

An Illustration with NSW Data Set*

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Abstract

In this paper, we study data issues of using matching estimators to estimate treatment effect. We first demonstrate that with proper data set, the matching assumptions can be justified for voluntary programs. Next we compare covariate matching and propensity score matching methods, and show that they do not dominate each other in term of data requirement. Finally we use the National Supported Work Demonstration data set to illustrate the issues discussed above.
I. Introduction


When the selection bias is only due to observables, matching is a useful tool to estimate treatment effect. The most attractive feature of matching, compared with the regression type estimators, such as that of Barnow et al (1980), is its non-parametric nature. Matching neither imposes functional form restrictions such as linearity nor assumes a homogeneous treatment effect in the population. Both assumptions are usually not justified either by economic theory or by the data. The first issue in this paper is to explore the plausibility of the assumptions for matching estimators. We argue that if proper variables, such as the information on the application indicator and the decision variables used by the program administration, are collected, the assumptions for matching estimators can be justified for voluntary programs.

Using covariate matching to correct the bias due to observables is intuitive, since the source of the bias is the difference of observables in the treated group and comparison group. Matching on covariates by definition will remove this difference and hence the
bias. When there are many covariates, it is impractical to match directly on covariates because of the curse of dimensionality. Taking the study of the Comprehensive Employment and Training Act by Westat (1981) as an example, for controlling only 12 covariates, the covariate matching scheme of Westat led to more than 6 million cells. Since the number of observations is far less than 6 million, most of cells are empty and it is very hard to find a good match on all 12 covariates. It is usually necessary to map the high dimension of covariates into a scalar through some metric, which measures the closeness of two observations. The most often used metric is Mahalanobis metric, e.g. Rubin (1980).

Another way to reduce the dimensionality is through propensity score matching. Rosenbaum and Rubin (1983) show that while covariate is the finest balancing score, propensity score is the coarsest balancing score. A balance score, \( b(x) \), is a function of the observed covariates such that the conditional distribution of \( x \) given \( b(x) \) is the same for treated and comparison groups, as defined in Rosenbaum and Rubin (1983). So matching on covariates and matching on the propensity score will both make the distribution of the covariates in the treated group the same as the distribution of the covariates in the comparison group.

Covariate matching faces the curse of dimensionality, and often encounters empty or small matching cells, while propensity score matching reduces the high dimension of covariates to a scalar, and can also balance the observables between treated group and comparison group. It is natural to ask whether propensity score matching needs less data (requires few observations) than covariate matching? This is the second issues considered in this paper.
The last issue in this paper is using the National Supported Work Demonstration (NSW) data set to illustrate the data issues of matching as well as to show that propensity score matching and covariate matching do not dominate each other. Their performance, like any other econometric evaluation method, crucially depends on the data set.

The remaining paper is organized as follows. Section 2 sets up the model using the potential outcome framework, Section 3 explores the plausibility of the assumptions for matching estimators, Section 4 studies data issues of covariate and propensity score matching methods, Section 5 is an illustration using the NSW data set, and Section 6 concludes the paper.

II. Model Setup

A fruitful framework for estimating treatment effects is the potential outcome framework dated back to Neyman (1923) and is widely used both in economics and statistics, such as, Roy (1951), Quandt (1972), Rubin (1974) and Holland (1986).

In the potential outcome framework, each individual has two potential outcomes \((Y_{0i}, Y_{1i})\) for a treatment, such as job training, education, or a welfare program. \(Y_{1i}\) is the outcome if individual \(i\) is treated and \(Y_{0i}\) is the outcome if individual \(i\) is not treated. Let \(T_i = 1\) indicate that individual \(i\) is treated and \(T_i = 0\) indicate otherwise. With \((Y_{0i}, Y_{1i})\) we can define different treatment effects, such as those in Heckman and Vytlacil (1999), as follows:

\[
\Delta_i = Y_{1i} - Y_{0i} \quad \text{Treatment Effect for Individual } i
\]

\[
\Delta_{ATE} = E[\Delta_i] \quad \text{Average Treatment Effect for the Population (ATE)}
\]

\[
\Delta_S = E[\Delta_i | i \in S] \quad \text{Average Treatment Effect for the Sub-Population } S
\]
When $S = \{ i: T_i = 1 \}$, $\Delta_s$ is the treatment effect on the treated (TT), denoted as $\Delta_{TT}$.

The average treatment effect at population (or sub-population) level can be estimated without bias either by experimental data or by observational data if the selection bias is only due to observables.

That the selection bias is only due to observables is formally characterized by the following two assumptions:

1. $M_{11}: (Y_0, Y_1) \perp T | X$  
   Conditional Independence Assumption

2. $M_{20}: 0 < \text{prob}(T = 1 | X) < 1$  
   Common Support Assumption

where $\perp$ is the notation for statistical independence as in Dawid (1979). $M_{-1}$ is also commonly referred as unconfoundedness assumption or exogeneity assumption.

Under $M_{-1}$ and $M_{-2}$

$$
\Delta_{TT} = E_{q(T=1)} \{ E[Y_1 | T = 1, X = x] - E[Y_0 | T = 1, X = x] \} \\
= E_{q(T=1)} \{ E[Y_1 | T = 1, X = x] - E[Y_0 | T = 0, X = x] \} \quad (1)
$$

Unbiased estimates of $E[Y_1 | T = 1, X = x]$ and $E[Y_0 | T = 0, X = x]$ can be obtained from the data and hence so can $\Delta_{TT}$. This is also true for $\Delta_{ATE}$ and for other $\Delta_s$.

Using the so-called balancing property:

$$
\text{prob}(X_i | T_i = 1, p(X_i) = p) = \text{prob}(X_i | T_i = 0, p(X_i) = p) = \text{prob}(X_i | p)
$$

Rosenbaum and Rubin (1983) prove that $M_{-1}$ and $M_{-2}$ imply

1. $P_{-1}: (Y_0, Y_1) \perp T | p(X)$, and

2. $P_{-2}: 0 < \text{prob}(T = 1 | p(X)) < 1$  

Following from $P_{-1}$ and $P_{-2}$:
\[ \Delta_{TT} = E_{p[T=1]} \{ E[Y_i | T=1, p(X) = p] - E[Y_0 | T=0, p(X) = p] \} \]
\[ = E_{p[T=1]} \{ E[Y_i | T=1, p(X) = p] - E[Y_0 | T=0, p(X) = p] \} \] (2)

Unbiased estimates of \( E[Y_i | T=1, p(X) = p] \) and \( E[Y_0 | T=0, p(X) = p] \) can also be obtained if \( p(X) \) is known. The advantage of formula (2) over formula (1) is that instead of controlling for a high-dimensional vector of \( X \), formula (2) only needs to control for a scalar \( p \).

III. The Plausibility of Matching Assumptions

Before going any further, an obvious question is whether the assumptions of \( M - 1 \) and \( M - 2 \) are plausible. Unfortunately in general there is no unambiguous answer to this question. Whether \( M - 1 \) and \( M - 2 \) are plausible or not needs to be argued case by case, and their plausibility depends on many factors, such as the richness of the data set, the nature of the treatment, the treatment effect under estimation, etc. The empirical evidence also suggests that there is no clear-cut answer for this question. Dehejia and Wahba (1999) successfully replicate the experimental benchmark by propensity score matching methods using the NSW data set. But Heckman et al (1998) reject the assumptions of matching estimators and assumptions of their extension, difference-in-difference matching, using the JTPA data set. They also find that selection bias due to observables is much larger than the one due to unobservables. Their work suggests that controlling for the bias due to observables is more important than controlling for the bias due to unobservables. Even if \( M - 1 \) and \( M - 2 \) are not justifiable and there is no prior knowledge on the magnitude of the bias due to unobservable and the bias due to observable, it is still useful to apply matching methods
to eliminate the bias due to observables first and then use different procedures to address
the bias due to unobservables.

Nevertheless $M - 1$ and $M - 2$ can be justified for a wide variety of applications
if a proper data set is available. In the United States as well as in many other countries,
like countries in the European Union, participation of the social programs is often
voluntary. For these programs, the treatment status is the result of two decisions: the
application decision made by each individual and the admission decision made by the
program administration. Let $A_i$ and $B_i$ be the indicators of these two decisions, so
$T_i = A_iB_i$. Under this scenario different treatment effects can be defined for the people
who apply for the program (i.e. for $i$ with $A_i = 1$):

\[
\Delta_{ATT-A} = [\Delta_i \mid A_i = 1] \quad \text{Average Treatment Effect for the Applicants}
\]

\[
\Delta_{TT-A} = E[\Delta_i \mid A_i = 1, T_i = 1] \quad \text{Treatment Effect on the Treated for Applicants}
\]

\[
\Delta_{UTT-A} = E[\Delta_i \mid A_i = 1, T_i = 0] \quad \text{Treatment Effect on the Untreated for Applicants}
\]

Essentially these are the treatment effects we are interested in,\(^1\) except in some
special cases such as if we try to evaluate what will happen if a program is expanded to
cover the whole population and is changed from a voluntary program into a mandatory
program. In that case we also need to know the treatment effect for the non-applicants.
These treatment effects for the applicants can be used to answer many interesting
questions, for instance, can the benefit of a program cover its cost? What will happen if
the coverage of a program is expanded?

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\(^1\) For the non-applicants, since they never participate in the program, their responses to the program have
little policy interest.
$M - 1$ and $M - 2$ for these treatment effects for the applicants can be justified if $A_i$ and $B_i$ can be observed separately. Taking $\Delta_{TT-A}$, treatment effect on the treated for applicants, as an example, it can be written as:\(^2\)

$$
\Delta_{TT-A} = E[\Delta_i | A_i = 1, T_i = 1]
= E[Y_{1i} - Y_{0i} | A_i = 1, B_i = 1]
= \{E[Y_{1i} | A_i = 1, B_i = 1] - E[Y_{0i} | A_i = 1, B_i = 0]\}
+ \{E[Y_{0i} | A_i = 1, B_i = 0] - E[Y_{0i} | A_i = 1, B_i = 1]\}
$$

First, $\Delta_{TT-A} = \Delta_{TT}$, i.e. the treatment effect on the treated for the applicants is the same as the treatment effect on the treated for the whole population, since both of them are the average treatment effect for the same group of people.\(^3\) Second, the first term $\{E[Y_{1i} | A_i = 1, B_i = 1] - E[Y_{0i} | A_i = 1, B_i = 0]\}$ is directly estimable from the data. The second term $\{E[Y_{0i} | A_i = 1, B_i = 0] - E[Y_{0i} | A_i = 1, B_i = 1]\}$ is the bias term and is due to observables if the program administration decision variables are collected, i.e. the program decision indicator, $B_i$, is independent of $Y_{0i}$ conditioning on the decision variables of $B_i$. When applying propensity score matching methods, we need to decide which $X$ should be included in the propensity score so $P-1$ will be satisfied and what functional form the propensity score should have. The balancing test in Dehejia and Wahba (1999) is useful to find the functional form of the propensity score but cannot tell which $X$ should be used, see Smith and Todd (2003). If we have sufficient knowledge of the

\(^2\) $\Delta_{UTT-A}$ can be justified in the same manner and $\Delta_{ATT-A}$ can be written as a weighted average of $\Delta_{TT-A}$ and $\Delta_{UTT-A}$.

\(^3\) Generally $\Delta_{ATT-A} \neq \Delta_{ATE}$ and $\Delta_{UTT-A} \neq \Delta_{UTT}$, where $UTT = \{i : T_i = 0\}$. 

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program and restrict our attention on the applicants, we need only include the $X$ used by the program administrator to select the applicants into the treatment.

The above argument relies on the assumption that we have the information on $A_i$ and $B_i$. Instead of focusing on devising different kinds of estimators based on ultimately untestable assumption(s), a feasible and more reliable alternative is to collect good data.\footnote{See Moffitt (1991) and Heckman et al (1998) on the importance of good (rich) data from other perspectives.} The application status indicator $A_i$ should be (but unfortunately has not been) included in many data sets, such as the Survey of Income and Program Participation (SIPP).

IV. **Data Issues of Covariate Matching and Propensity Matching Estimators**

The common approaches to control for the bias due to observable variables in the matching literature include matching on covariates or on the propensity score, sub-classification by covariates or by the propensity score, and weighting by the propensity score. Imbens (2003) provides an excellent survey. We will focus on one-to-one matching estimators since one-to-one matching estimators are widely used in the empirical studies and it is important to understand their properties. One-to-one matching involves selecting a single observation from the comparison sample to match each observation in the treated sample by some metric. Though matching on covariates or on the propensity score can both remove the bias due to observables, if there are many covariates, especially continuous ones, matching on covariates runs into the curse of dimensionality. Since the work of Rosenbaum and Rubin (1983), propensity score matching has dominated the literature on matching. In most cases, it is easier to find observations with similar values of propensity score than with similar values of covariates, as argued in Rosenbaum (1995). Does this mean that propensity score matching requires fewer observations than...
covariate matching? In order to answer this question, we need to examine more closely
how covariate matching and propensity score matching work.

Define the two potential outcome equations and the selection equation as:

\[ Y_{it} = f_t(X_i) + \epsilon_{it}, \]  
\[ \epsilon_{it} \text{ is iid with } E[\epsilon_{it} | X_i] = 0; \]
\[ Y_{0i} = f_0(X_i) + \epsilon_{0i}, \]  
\[ \epsilon_{0i} \text{ is iid with } E[\epsilon_{0i} | X_i] = 0; \]
\[ T_i = I(T_i^* > 0), \]  
\[ I(\cdot) \text{ is the indicator function; and} \]
\[ T^* = h(X_i) + \mu_i, \]  
\[ \mu_i \text{ is iid with } E[\mu_i | X_i] = 0 \text{ and } CDF \ G(\cdot). \]

The basic ideas of covariate matching are:

(1) \[ X_i = X_j \Rightarrow f_t(X_i) = f_t(X_j), \]  
\[ t = 0, 1; \]  
and

(2) \[ d(X_i, X_j) < \varepsilon \Rightarrow d'(f_t(X_i), f_t(X_j)) < \delta, \]  
\[ t = 0, 1, \]  
where \(d\) and \(d'\) are some metrics in mathematical sense.

Assumption (1) justifies exact matching. Assumption (2) means that \(f_t\) is continuous at \(X\) and it justifies neighborhood matching.

For simplicity, we assume that exact matching is possible.\(^5\) Through covariate matching, the observation \(i\) in the treated sample is matched with the observation \(j\) in the comparison sample if \(X_i = X_j = x\). Define:

\[ \tilde{\Delta}_{it}^c = Y_{it} - Y_{0j}, \]
\[ = f_t(X_i) + \epsilon_{it} - f_0(X_j) - \epsilon_{0j}, \]
\[ = \{f_t(x) + \epsilon_{it} - [f_0(x) + \epsilon_{0i}]\} + \{\epsilon_{0i} - \epsilon_{0j}\} \]
\[ = \Delta_i + \{\epsilon_{0i} - \epsilon_{0j}\} \]

\(^5\) When exact matching is impossible, the discussions are still approximately true if we can match on some sufficiently small neighborhood of \(X\).
where $\Delta_i$ is the true treatment effect for individual $i$. Denote $m(x)$ as the number of matching pairs in an $x-$cell which have the same covariate $x$; then the average treatment effect at $x$ can be estimated by:

$$\hat{\Delta}^C(x) = \frac{1}{m(x)} \sum_{i=1}^{m(x)} \Delta_i$$

$$= \frac{1}{m(x)} \sum_{i=1}^{m(x)} \Delta_i + \frac{1}{m(x)} \sum_{i=1}^{m(x)} \varepsilon_{0i} - \frac{1}{m(x)} \sum_{j=1}^{m(x)} \varepsilon_{0j}$$

$$= \Delta(x) + \frac{1}{m(x)} \sum_{i=1}^{m(x)} \varepsilon_{0i} - \frac{1}{m(x)} \sum_{j=1}^{m(x)} \varepsilon_{0j}$$

where $\Delta(x)$ is the average treatment effect at $x$. It is clear that $\hat{\Delta}^C(x)$ is an unbiased and consistent estimator of the average treatment effect at $x$. Let $r^C$ be the number of covariate matching cells, and let $n^C = \sum_{r^C} m(x)$ be the total number of matched pairs in the whole sample ($n^C$ also equals the number of observations in the treated sample). The treatment effect on the treated can be estimated by the following estimator:

$$\hat{\Delta}^C_{TT} = \frac{1}{n^C} \sum_{i=1}^{n^C} \Delta_i$$

$$= \frac{1}{n^C} \sum_{i=1}^{n^C} \Delta(x) + \frac{1}{n^C} \sum_{i=1}^{n^C} \varepsilon_{0i} - \frac{1}{n^C} \sum_{j=1}^{n^C} \varepsilon_{0j}$$

$$= \Delta_{TT} + \frac{1}{n^C} \sum_{i=1}^{n^C} \varepsilon_{0i} - \frac{1}{n^C} \sum_{j=1}^{n^C} \varepsilon_{0j}$$

The closeness of the covariates of each matching pair plays a crucial role in the covariate matching and itself is enough to guarantee the reliability of the estimator under the assumptions of $M \geq 1$ and $M \geq 2$ and the continuity of $f_i$. 

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The theory behind propensity score matching is quite different from covariate matching.

The basic ideas of propensity score matching are:

(1) \( \text{prob}(X_i \mid T_i = 1, p(X_i) = p) = \text{prob}(X_i \mid T_i = 0, p(X_i) = p) = \text{prob}(X_i \mid p) \), the balancing property; and

(2) \( d(p_k, p_t) < \varepsilon \Rightarrow d'(\text{prob}(X_i \mid p_k), \text{prob}(X_j \mid p_t)) < \delta \)

These two ideas are parallel to the two ideas of covariate matching. Assumption (1) says that when the matching is exact at the propensity score \( p \), then the distribution of \( X \) will be the same for the treated sample and the comparison sample at \( p \). Assumption (2) says if exact matching is impossible and instead matching is on some neighborhood of \( p \), the distribution of \( X \) is still approximately the same for the treated sample and the comparison sample within the neighborhood of \( p \).

In the propensity score matching methods, the observation \( i \) in the treated sample is matched with the observation \( j \) in the comparison sample if \( p(X_i) = p(X_j) = p \).

Define:

\[
\hat{\Delta}_i^p = Y_i - Y_{0j} = f_i(X_i) + \varepsilon_i - \{f_0(X_j) + \varepsilon_{0j}\} = \{f_i(X_i) + \varepsilon_i - [f_0(X_i) + \varepsilon_{0i}]\} + \{f_0(X_i) - f_0(X_j) + \varepsilon_{0i} - \varepsilon_{0j}\} = \Delta_i + \{f_0(X_i) - f_0(X_j) + \varepsilon_{0i} - \varepsilon_{0j}\}
\]
Using $\hat{\Delta}^p_i$ as the building block, and denoting $m(p)$ as the number of matching pairs in a $p$–cell which has the same propensity score $p$, we can estimate the average treatment effect at $p$ by:

$$\hat{\Delta}^p (p) = \frac{1}{m(p)} \sum_{i=1}^{m(p)} \Delta_i,$$

$$= \frac{1}{m(p)} \sum_{i=1}^{m(p)} \Delta_i + \frac{1}{m(p)} \sum_{i=1}^{m(p)} e_{0,i} - \frac{1}{m(p)} \sum_{j=1}^{m(p)} e_{0,j} + \frac{1}{m(p)} \sum_{i=1}^{m(p)} f_0(x_i) - \frac{1}{m(p)} \sum_{j=1}^{m(p)} f_0(x_j),$$

where $\Delta(p)$ is the average treatment effect at $p$. $\hat{\Delta}^p (p)$ is an unbiased and consistent estimator of the average treatment effect at $p$.

Let $r^p$ be the number of propensity score matching cells, and $n^p = \sum_{i=1}^{r^p} m(p)$ be the number of matched pairs in the whole sample ($n^p$ also equals the number of observations in the treated sample). We can estimate the treatment effect on the treated by the following estimator, which is widely used in the matching literature:

$$\hat{\Delta}^p_{TT} = \frac{1}{n^p} \sum_{i=1}^{n^p} \hat{\Delta}_i^p,$$

$$= \frac{1}{n^p} \sum_{i=1}^{n^p} \Delta_i + \frac{1}{n^p} \sum_{i=1}^{n^p} e_{0,i} - \frac{1}{n^p} \sum_{j=1}^{n^p} e_{0,j} + \frac{1}{n^p} \sum_{i=1}^{n^p} f_0(x_i) - \frac{1}{n^p} \sum_{j=1}^{n^p} f_0(x_j),$$

$$= \Delta_{TT} + \left\{ \frac{1}{n^p} \sum_{i=1}^{n^p} e_{0,i} - \frac{1}{n^p} \sum_{j=1}^{n^p} e_{0,j} \right\} + \left\{ \frac{1}{n^p} \sum_{i=1}^{n^p} f_0(x_i) - \frac{1}{n^p} \sum_{j=1}^{n^p} f_0(x_j) \right\}.$$
It is clear that $\hat{\Delta}_{TT}$ is also an unbiased estimator for $\Delta_{TT}$. The second term, 
\[
\left\{ \frac{1}{n^p} \sum_{i=1}^{n^p} e_{0i} - \frac{1}{n^p} \sum_{j=1}^{n^p} e_{0j} \right\},
\]
will go to zero as sample size goes to infinity. The third term, 
\[
\left\{ \frac{1}{n^p} \sum_{i=1}^{n^p} f_0(x_i) - \frac{1}{n^p} \sum_{j=1}^{n^p} f_0(x_j) \right\},
\]
needs to be balanced out.

It is very possible that individuals with the same propensity score will have very different treatment outcomes, i.e. $p$ approximately the same does not imply $X$ hence doesn't imply treatment outcome, $f(X)$, approximately the same. Because of the balancing property this will not be a problem if the number of observations at each propensity score is large. This can be easily seen if we compare propensity score matching methods to a randomized experiment. The foundation of a randomized experiment is $\frac{\text{prob}(X, \nu | \text{treated})}{\text{prob}(X, \nu | \text{control})} \equiv \frac{\nu}{\nu}$ where $X$ is observable and $\nu$ is unobservable. The balancing property plays a similar role in propensity score matching, but propensity score matching methods differ from randomization in two important ways. First, a randomized experiment balances the distributions of both observables and unobservables between treated and comparison samples, but propensity score matching only balances the observables. This is why the independence assumption $M - 1$ is needed. Second, a randomized experiment balances the distributions for the whole sample, but propensity score matching balances the distributions at each individual propensity score value. In other words, under $M - 1$ and $M - 2$, the matched sample at each propensity score value $p$ is equivalent to a randomized sample. The estimate of propensity score matching can be thought as a weighted average of the estimates from many mini randomized experiments (at different $p$'s). The overall quality of the estimation depends on the quality of each of these mini randomized experiments. A
substantial sample size is needed to obtain a meaningful estimate from a randomized experiment and this is translated into a sufficiently large sample size at each $p$ for a meaningful propensity score matching estimate.

When comparing covariate matching with propensity score matching, we note that the advantage of propensity score matching over covariate matching is often characterized by dimensionality reduction, which composes two aspects. One aspect of dimensionality reduction is that instead of controlling high-dimension $X$, controlling the propensity score $p$, a scalar, is enough. Nonetheless the data requirement we discussed is related to the other aspect of dimensionality reduction, namely in general the number of $p$-cells, $r^p$ is less than the number of $X$-cells, $r^C$ (also see the discussion in Angrist and Hahn 1999). Let us consider two polar cases. One polar case is a randomized experiment. This is the strongest case for the propensity score matching. Since $p(X_i)$ is the same for every individual in the randomization, $r^p$ is 1. The dramatic reduction of the data requirement for the randomized experiment is the result of the drastic reduction of $r^p$ compared with $r^C$. The other polar case is that in which the correspondence between $p$ and $X$ is one-to-one. In this case if exact matching is possible, then matching on the propensity score or on covariates is equivalent and both require same amount of data since in this case people with same $X$ must have same $p$, and vice versa. If exact matching is impossible and instead we match on some neighborhood of the propensity score, the story is different. We note that it is a fact there does not exist a one-to-one and bi-continuous (i.e. both the function and its inverse function are continuous) correspondence between $R^n$ space and $R^1$ space, i.e. $R^n$ space and $R^1$ space are not a homoeomorphism. It is natural to assume that $p(X)$ is a continuous function of $X$ and this implies that
$p^{-1}(X)$ is not a continuous function of $p$. The implication of this mathematical fact is shown in Figure 1. On the one hand, if $X$'s, like $X_1$ and $X_2$, lie in the set $A$, then their $p(X)$'s, i.e., $p_1$ and $p_2$, must lie in the set $B$ (this follows from the continuity of $p(X)$). On the other hand, there must be always some $X$'s, e.g., $X_3$ and $X_4$, that lie outside the set $A$, but whose $p(X)$'s, i.e., $p_3$ and $p_4$, are in the set $B$ (this follows from the discontinuity of $p^{-1}(X)$). Their corresponding treatment outcomes can be quite different from the ones in the set $A$. Matching by the propensity score on some neighborhood of the propensity score has the risk of matching $p_1$ with $p_4$, whose $f(X_1)$ and $f(X_4)$ are quite different even though their propensity scores are similar and the correspondence between $X$ and $p$ is one to one. To average this kind of mismatching out, propensity score matching must rely on the balancing property and needs the neighborhood of $p$ to contain a relatively large number of observations. In this case, the advantage of matching on covariates is obvious.

Whether propensity score matching needs less data hinges on how large the difference between $r^p$ and $r^c$ is. Briefly, the reduction of the data requirement of propensity score matching relies on the reduction of the cell number. The reduction of the cell number creates the risk of mismatching. To average out the risk of mismatching requires a large cell size $m(p)$. The combination of $r^p$, $r^c$ and $m(p)$ ultimately determines the relative data requirements between propensity score matching and covariate matching. Propensity score matching and covariate matching do not dominate each other regarding the data requirement.
V. An Illustration with the National Supported Work Demonstration Data Set

The NSW Demonstration is a randomized experiment conducted from 1975 to 1980 to estimate the effects of a “supported” work experience on the disadvantaged population, such AFDC recipients and ex-offenders. This experiment has 3,214 observations in the treated sample and 3,402 in the control sample. The NSW data set has played an important role in the treatment effect literature. Lalonde (1986) uses a subset of the NSW data set, combined with the PSID and the CPS data, to evaluate different non-experimental estimators. He uses the estimate from the NSW data set as the benchmark, then drops the control group in the NSW data and constructs other comparison groups from the CPS and the PSID. Different estimators have been applied to the constructed data sets. Different estimators have produced very different estimates and often have failed to replicate the benchmark. Fraker and Maynard (1987) use a similar approach but emphasize the sensitivity of estimates to the selection of the comparison groups from the NSW and the CPS data, and they reach a similar conclusion as LaLonde. As a response, Heckman and Hotz (1989) devise tests to aid the choice among estimators. Using propensity score methods, Dehejia and Wahba (1999) successfully replicate the benchmark result, but Smith and Todd (2003) suggest that the success of propensity score matching methods in Dehejia and Wahba (1999) has something to do with the data selected by Dehejia and Wahba instead of the propensity score matching method, per se.6

The data sets used in LaLonde (1986), Dehejia and Wahba (1999) and Smith and Todd (2003) are different. The Dehejia and Wahba data set is a subset of LaLonde data set and it excludes the observations with missing earnings variable in 1974. Smith and

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6 The data set in Heckman and Hotz (1989) is different from the one in LaLonde (1986), though both are subsets of the NSW data set.
Todd data set is a subset of Dehejia and Wahba data set and it excludes the observations that were randomized after April of 1976 and Smith and Todd argues that including these observations is problematic.

Similar to their work, we apply propensity score matching methods, Mahalanobis metric, and other two matching metrics proposed in Zhao (2003) and discussed in Imbens (2003) to both the LaLonde data set and the Dehejia and Wahba data set.

The first metric considered in Zhao (2003) and Imbens (2003) is as follows: let the propensity score \( p(X) = G(X\beta') \), and consider the following metric:

\[
d_1 = \sum_{k=1}^{K} |X_{ki} - X_{kj}| \cdot |\beta_k|
\]

This metric incorporates information on both \( X \) and \( p \), and weights each coordinate of \( X \) by its marginal effect on the propensity score.\(^7\)

The second metric is incorporated outcome information. Assume \((Y_{0i}, Y_{1i})\) and \( X \) have linear relationships, such that

\[
Y_{it} = f_{it}(X_i) + \varepsilon_{it} = X_i\alpha_i + \varepsilon_{it}, \ t = 0, 1
\]

Define the metric as:

\[
d_2 = \sum_{k=1}^{K} |X_{ki} - X_{kj}| \cdot |\alpha_{k}\|
\]

This metric weights the coordinates of \( X \) by their marginal effects to the potential outcomes. It is a natural measurement for closeness of two observations in term of their potential outcomes.

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\(^7\) Strictly speaking, \( \beta \) can be interpreted as the marginal effect only if it is estimated from the LPM. For other models, like probit and logit, though \( \beta \) is not the marginal effect but it is still proportioned to the marginal effect.
We refer matching by metrics $d_1$ and $d_2$ as covariate & propensity score matching and covariate & outcome matching, respectively. All treatment effects are estimated by one to one matching. Since the CPS is more representative than the PSID and since we want to examine the effectiveness of different matching methods and do not want other sample selection procedures to contaminate the matching process, our estimation is focused on the whole CPS sample, i.e. CPS-SSA-1 in LaLonde (1986) and CPS-1 in Dehejia and Wahba (1999).

Table 1 shows results from different matching metrics using the Dehejia and Wahba data set. The propensity score specification in Table 1 is the same as the specification in Dehejia and Wahba (1999). Measured by the closeness to the benchmark, results from different metrics are very similar and there is no evidence that one estimator dominates the other estimator. Matching without replacement performs more poorly than matching with replacement, which is consistent with Dehejia and Wahba (1999). Imposing the common support condition has little effect on the results. For some estimators it increases the bias, and for others it reduces the bias, but these changes are small. This is not surprising. Unlike sub-classification estimators, one-to-one matching estimators automatically solve the common support problem.

As discussed earlier, the estimate from propensity score matching is the weighted average of the estimates at different propensity score values. The overall quality of the estimation relies on the quality of estimation at each propensity score value. Besides the final estimate, it is also interesting to examine more closely the intermediate estimates. Figure 2 shows the propensity scores of the matched pairs. They are very well matched in terms of the propensity score. Figure 3 is the treatment effect estimated at a pair level. It
highlights that people with similar propensity score can have very different treatment effects. Using age as example, Figure 4 shows that people with a very similar propensity score value can have very different covariates. As discussed in Section 4, the difference of covariates at pair level does not matter as long as the averages of covariates are similar at cell level. We stratify the matched pairs into 18 cells by the propensity score of the treated observation in each pair. The width of each cell is 0.05 (since there is no treated observation with propensity score value larger than 0.9, there are 18 cells). Figure 5 shows the treatment effects for each cell estimated from the NSW experiment and propensity score matching. There is less volatility than at the pair level, but they are still very noisy. Contrary to the common intuition that the people have higher propensity score values also have larger treatment effects, it seems that the treatment effect is independent of the propensity score. The independence of the treatment effect and the propensity score partially explains why matching methods are successful in the Dehejia and Wahba (1999). Table 2 shows that the means of the covariates of the treated sample and the comparison sample in each cell. The balance of the covariates in each cell is a necessary condition for the propensity score matching methods to work. Table 2 shows some cells and some covariates in certain cells are indeed balanced, but the majority of them are not. Dehejia and Wahba (1999) devised a test to make the covariates balanced in each cell before carrying out matching, which is different from the issue we discuss here. Even if the covariates are balanced before matching, they could become unbalanced after matching, but the unbalance will become unlikely as the sample size increases.

The estimates from the LaLonde data set are shown in Table 3. Contrary to the estimates from the Dehejia and Wahba data set, all methods fail to replicate the
benchmark from the NSW experiment and most estimates even do not have right sign. The estimation from OLS is as good as the estimations from propensity score matching and the majority of the estimations from covariate matching, if not better.

The only difference of the LaLonde data set from the Dehejia and Wahba data set is that the Dehejia and Wahba data set has information on the earnings in 1974 (two years prior the treatment) but the LaLonde data set does not. It is hard to imagine that this difference is responsible for the failure of the matching estimators in the LaLonde data set, though the importance of pre-program earning history in the program evaluation is well known since the discovery of the famous Ashenfelter dip in Ashenfelter (1978). In order to explore this issue further, we pretend that we do not have the 1974 earning information in Dehejia and Wahba data set and estimate the treatment effects without using the earning variable of 1974. The results are reported in Table 4. Compared Table 1 with Table 4, it can be seen that the contribution of the 1974 earning history in improving the estimation of the treatment effects is marginal at most. With or without the 1974 earning history, the estimates from Dehejia and Wahba data set are closed to the NSW experimental benchmark.

VI. Conclusions

Selection bias due only to observables is a strong assumption, but for voluntary programs if we have the data on the application indicator and on the variables used by the program administrator to make the selection decision, we can justify this assumption.

With a proper data set and if the selection on observables assumption is justifiable, matching methods are useful tools to estimate treatment effects. There is no clear winner among matching estimators considered here. Propensity score matching
methods rely on the balancing property and require a large number of observations at each propensity score value, which is not required by covariate matching methods, while covariate matching methods face the curse of dimensionality and often encounter small cell or empty cell problem, so they do not dominate each other in term of data requirement in finite samples.

A major data requirement for propensity score matching is that at each propensity score value the number of observations is large. If this condition fails, it could affect the final results. It is important to check whether the covariates are balanced after matching at each propensity score value.

The failure of matching methods in the LaLonde data set highlights that, like any non-experimental estimator, the behavior of matching estimators largely depends on the data structure at hand. Matching is a useful estimator under suitable conditions, but it is definitely not the estimator for every evaluation. There is no easy way out in social program evaluation. A successful evaluation study requires detailed knowledge of the program, a good data set, and a careful consideration and choice of the estimation strategy.
Reference


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### Table 1 Estimates from Various Matching Estimators Using Dehejia and Wahba Data

#### Panel A: Matching with Replacement

<table>
<thead>
<tr>
<th>Methods</th>
<th>With Common Support Condition</th>
<th>Without Common Support Condition</th>
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<td>Treatment Effect</td>
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#### Panel B: Matching without Replacement

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Note: 1. The specification of the propensity score is the same as in Dehejia and Wahba (1999), including age, age squared, education, education squared, no degree, married, black, hispanic, re74, re75, u74, u75, education*re74 and age cubed.
2. The specification of the OLS is the same as the specification of the propensity score.
3. The outcome equation in the covariate & outcome matching is estimated using only the treated data by OLS and without any higher order and interaction term.
Table 2 The Covariate Means in Each Cell after Propensity Score Matching Using Dehejia and Wahba Data

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<th>Education_Treated</th>
<th>Education_Control</th>
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Table 3: Estimates from Various Matching Estimators Using Lalonde Data

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Panel B: Matching without Replacement

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<td>Propensity Score Matching (LPM)</td>
<td>-1537.579</td>
<td>-2423.8827</td>
</tr>
<tr>
<td>Propensity Score Matching (Weighted LPM)</td>
<td>-1145.6929</td>
<td>-2031.9866</td>
</tr>
<tr>
<td>Covariate Matching (Mahalanobis)</td>
<td>-761.2005</td>
<td>-1647.5042</td>
</tr>
<tr>
<td>Covariate &amp; Propensity Score (Probit)</td>
<td>-1824.3922</td>
<td>-2710.6961</td>
</tr>
<tr>
<td>Covariate &amp; Propensity Score (Logit)</td>
<td>-1801.1467</td>
<td>-2686.4504</td>
</tr>
<tr>
<td>Covariate &amp; Propensity Score (LPM)</td>
<td>-2429.0461</td>
<td>-3315.3498</td>
</tr>
<tr>
<td>Covariate &amp; Propensity Score (Weighted LPM)</td>
<td>-2224.2692</td>
<td>-3110.5729</td>
</tr>
</tbody>
</table>

Note: 1. The specification of the propensity score is the same as in Dehejia and Wahba (1999), including age, age squared, education, education squared, no degree, married, black, hispanic, re75, u75 and age cubed.
2. The specification of the OLS is the same as the specification of the propensity score.
3. The outcome equation in the covariate & outcome matching is estimated using only the treated data by OLS and without any higher order and interaction term.
### Table 4 Estimates from Various Matching Estimators Using Dehejia and Wahba Data without 1974 Earning History

#### Panel A: Matching with Replacement

<table>
<thead>
<tr>
<th>Methods</th>
<th>With Common Support Condition</th>
<th>Without Common Support Condition</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Treatment Effect</td>
<td>Bias</td>
</tr>
<tr>
<td>NSW Experiment (Benchmark)</td>
<td>1794.3424</td>
<td>0</td>
</tr>
<tr>
<td>Simple Mean Difference</td>
<td>-1542.2862</td>
<td>-3336.6286</td>
</tr>
<tr>
<td>OLS</td>
<td>947.8409</td>
<td>-846.5015</td>
</tr>
<tr>
<td>Propensity Score Matching (Probit)</td>
<td>1742.4868</td>
<td>-51.8556</td>
</tr>
<tr>
<td>Propensity Score Matching (Logit)</td>
<td>1062.9881</td>
<td>-731.3543</td>
</tr>
<tr>
<td>Propensity Score Matching (LPM)</td>
<td>-18.6278</td>
<td>-1812.9702</td>
</tr>
<tr>
<td>Propensity Score Matching (Weighted LPM)</td>
<td>889.7427</td>
<td>-904.5997</td>
</tr>
<tr>
<td>Covariate Matching (Mahalanobis)</td>
<td>1379.3894</td>
<td>-414.953</td>
</tr>
<tr>
<td>Covariate &amp; Propensity Score (Probit)</td>
<td>1228.9009</td>
<td>-565.4415</td>
</tr>
<tr>
<td>Covariate &amp; Propensity Score (Logit)</td>
<td>1837.8201</td>
<td>43.4777</td>
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<tr>
<td>Covariate &amp; Propensity Score (LPM)</td>
<td>1353.0539</td>
<td>-441.2885</td>
</tr>
<tr>
<td>Covariate &amp; Propensity Score (Weighted LPM)</td>
<td>1473.6523</td>
<td>-320.6901</td>
</tr>
<tr>
<td>Covariate &amp; Outcome</td>
<td>1848.228</td>
<td>53.8566</td>
</tr>
</tbody>
</table>

#### Panel B: Matching without Replacement

<table>
<thead>
<tr>
<th>Methods</th>
<th>With Common Support Condition</th>
<th>Without Common Support Condition</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Treatment Effect</td>
<td>Bias</td>
</tr>
<tr>
<td>Propensity Score Matching (Probit)</td>
<td>1875.3385</td>
<td>80.9961</td>
</tr>
<tr>
<td>Propensity Score Matching (Logit)</td>
<td>1296.0695</td>
<td>-498.2729</td>
</tr>
<tr>
<td>Propensity Score Matching (LPM)</td>
<td>-785.504</td>
<td>-2579.8464</td>
</tr>
<tr>
<td>Propensity Score Matching (Weighted LPM)</td>
<td>848.9815</td>
<td>-945.3609</td>
</tr>
<tr>
<td>Covariate Matching (Mahalanobis)</td>
<td>892.8058</td>
<td>-901.4366</td>
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<tr>
<td>Covariate &amp; Propensity Score (Probit)</td>
<td>1523.9989</td>
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<tr>
<td>Covariate &amp; Propensity Score (Logit)</td>
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</tr>
<tr>
<td>Covariate &amp; Outcome</td>
<td>883.0169</td>
<td>-911.3255</td>
</tr>
</tbody>
</table>

**Note:**
1. The specification of the propensity score is linear without any higher order and interaction term.
2. The specification of the OLS is the same as the specification of the propensity score.
3. The outcome equation in the covariate & outcome matching is estimated using only the treated data by OLS and without any higher order and interaction term.
Figure 1 Neighborhood Matching
With One to One Correspondence between $X$ and $p$
Note: 1. There are total 185 matched pairs;
   2. The matched pairs are sorted by the propensity score of the treated;
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   2. The matched pairs are sorted by the propensity score of the treated;
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2. The matched pairs are sorted by the propensity score of the treated;
Note: 1. There are total 185 matched pairs;
   2. The matched pairs are sorted by the propensity score of the treated;
   3. The width of the cell in Figure 5 is 0.05. Since there is no observation with propensity score larger than 0.9, there are only 18 cells. The numbers in the plot are the numbers of matched pairs in each cell.